

## ORIGINAL ARTICLE

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## Semi-automatic DNA profiling in a Hungarian Romany population using the STR loci HumVWFA31, HumTH01, HumTPOX, and HumCSF1PO

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**Abstract** A population study of Hungarian Romanies was carried out for the STR loci HumVWFA31, HumTH01, HumTPOX, and HumCSF1PO. After multiplex PCR amplification semi-automatic DNA profiling was performed using an ALF DNA sequencer. At the loci investigated there was little and no evidence for departures from Hardy-Weinberg expectations and linkage equilibrium, respectively. The allele sizing accuracy of the ALF DNA sequencer was increased to a high level (99.97% on average) by applying external and internal markers. Allele frequency distributions of the STR loci, with one exception, were significantly different between the Romany and other Hungarian population databases. On the other hand, however, only small differences in frequencies of individual phenotypes were found.

**Key words** PCR · Multiplex STR profiling · Automated DNA sequencer · Romany (Gypsy) population · Hungary

### Introduction

The high variability of the short tandem repeats (STRs) and their relative simplicity when used as polymorphic markers, makes them useful tools for population genetics and genetic identification purposes [1, 2].

Prior to introduction of STR characterization to forensic casework in Hungary, population surveys have been started in the relevant populations [3–5]. In Hungary the Romany population is the largest of the ethnic minorities representing about 6% of the Hungarian inhabitants. The

Romany population, an Orientalo-Indid group of Caucasians, most likely originated from the north-western part of India, and started to settle in great numbers in Hungary in the 15th century [6]. The Romanies formed genetically closed population groups throughout the country.

While Romany population data for numerous conventional blood type loci exist in Hungary [7, 8], there have not been such population data generated for forensically interesting DNA polymorphisms. The use of an ALF DNA sequencer and fluorescent quadruplex amplification allowed us to rapidly obtain the four-locus (HumVWFA31, HumTH01, HumTPOX, and HumCSF1PO [2, 9, 10]) STR profiles from a Romany population of Baranya county in Hungary. The sizing accuracy of the ALF DNA sequencer was also evaluated since relatively little data have been available for this instrument [11] as compared to other automated DNA sequencers. Furthermore, the Romany population database established here was compared with other Hungarian population databases.

### Materials and methods

#### Population sample

The sample consisted of 135 unrelated Hungarian Romany ("Roma") individuals residing in Baranya county (south-western Hungary).

#### Multiplex amplification conditions and detection system

The coamplification of HumVWFA31, HumTH01, HumTPOX and HumCSF1PO loci was performed using reagents provided in the GenePrint Fluorescent Quadruplex STR System CSF1PO-TPOX-TH01-vWA (Promega, Madison, Wis.) according to the manufacturer's instructions. The PCR products were analysed on an ALF DNA sequencer (Pharmacia) using standard gel format (19 cm well-to-read distance) as previously described [12]. The fragment size of the amplified products was determined with the Promega allelic ladders loaded in lanes 3, 15, 27, and 38. Two fluorescent internal markers were used in each lane to compensate for any mobility shift between the lanes. A 100 bp internal marker was purchased from Pharmacia, and a 271 bp fragment (allele 9) of the HumHPRTB STR locus was obtained after amplification accord-

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ing to Szibor et al. [13], where the reverse primer was fluorescently labeled. The measurement error of computer-generated allele sizing was determined for the Promega allelic ladders and individual samples. The accuracy was calculated for each allele using the formula  $\% \text{ accuracy} = (1 - |\text{estimated size} - \text{expected size}| / \text{expected size}) \times 100$  [11].

Statistical methods

Possible divergence from Hardy-Weinberg expectations (HWE) was determined by calculating the unbiased estimate of the expected homozygote/heterozygote frequencies [14] and the exact test [15]. The interclass correlation [16] and  $s_k^2$  criteria [17] were used to test for allelic independence between/among STR loci. Population homogeneity was examined using a computerized G-statistic test of  $2 \times C$  contingency tables. Interpopulation differences were evaluated by calculating the mean phenotype difference values (MVs) [18] and by a scatter plot comparison [19]. In the MV investigation 100 individuals were randomly selected out of each population sample. In the scatter plot analysis we pooled all individual profiles from the databases to be compared. The likelihood values (LR) estimates of the four-locus profiles were computed multiplying the allele frequencies from each population. The respective LRs of every target profile were then plotted on a logarithmic scale (i.e. a scatter plot). Each axis of the plot assigned from which population database a LR log was estimated.

Results and discussion

Allele frequencies for the four STR loci in the Romany population sample are shown in Table 1. All alleles differed in size by one repeat unit (i.e. 4 base pairs) for all loci, except for the HumTH01 allele 9.3. All the loci did not deviate from HWE based on the exact test (Table 1). The excess of homozygous HumVWFA31 types reached the significance level of 0.03. However, no particular homozygous type was predominantly responsible for this excess (Table 2). An explanation for the surplus is that a

**Table 1** Allele frequencies and HWE test for four STR loci in 135 unrelated Hungarian Romany individuals

| Allele                  | VWA   | TH01  | TPOX  | CSF1PO |
|-------------------------|-------|-------|-------|--------|
| 6                       |       | 0.237 |       |        |
| 7                       |       | 0.085 |       | 0.004  |
| 8                       |       | 0.233 | 0.433 | 0.019  |
| 9                       |       | 0.181 | 0.067 | 0.011  |
| 9.3                     |       | 0.259 |       |        |
| 10                      |       | 0.004 | 0.144 | 0.289  |
| 11                      |       |       | 0.352 | 0.252  |
| 12                      |       |       | 0.004 | 0.381  |
| 13                      |       |       |       | 0.041  |
| 14                      | 0.085 |       |       | 0.004  |
| 15                      | 0.070 |       |       |        |
| 16                      | 0.333 |       |       |        |
| 17                      | 0.252 |       |       |        |
| 18                      | 0.233 |       |       |        |
| 19                      | 0.019 |       |       |        |
| 20                      | 0.007 |       |       |        |
| Exact test <sup>a</sup> | 0.296 | 0.856 | 0.083 | 0.070  |

<sup>a</sup>These values are probability values

**Table 2** Observed and expected homozygous VWA types in 135 Hungarian Romany individuals

| Homozygous type | Observed | Expected |
|-----------------|----------|----------|
| 14–14           | 2        | 0.98     |
| 15–15           | 2        | 0.67     |
| 16–16           | 15       | 15.00    |
| 17–17           | 13       | 8.56     |
| 18–18           | 11       | 7.35     |
| 19–19           | 0        | 0.05     |
| 20–20           | 0        | 0.01     |
| Homozygosity    | 0.319    | 0.239    |

Homozygosity test:  $P = 0.030$

**Table 3** Standard deviation (SD) of estimated size and maximum size range for the STR alleles obtained by semi-automatic DNA typing of 135 Hungarian Romany individuals<sup>a</sup>

| Locus  | Allele | SD (bp) | Range (bp) | N  |
|--------|--------|---------|------------|----|
| VWA    | 14     | 0.07    | 0.3        | 21 |
|        | 15     | 0.05    | 0.2        | 17 |
|        | 16     | 0.06    | 0.3        | 75 |
|        | 17     | 0.07    | 0.3        | 55 |
|        | 18     | 0.07    | 0.3        | 52 |
|        | 19     | 0.04    | 0.1        | 5  |
| TH01   | 6      | 0.06    | 0.2        | 55 |
|        | 7      | 0.06    | 0.2        | 23 |
|        | 8      | 0.05    | 0.2        | 53 |
|        | 9      | 0.06    | 0.2        | 46 |
|        | 9.3    | 0.06    | 0.3        | 61 |
| TPOX   | 8      | 0.08    | 0.4        | 96 |
|        | 9      | 0.08    | 0.2        | 16 |
|        | 10     | 0.05    | 0.2        | 39 |
|        | 11     | 0.06    | 0.2        | 77 |
| CSF1PO | 8      | 0.08    | 0.2        | 5  |
|        | 9      | 0.10    | 0.2        | 3  |
|        | 10     | 0.10    | 0.4        | 68 |
|        | 11     | 0.09    | 0.4        | 65 |
|        | 12     | 0.11    | 0.6        | 79 |
|        | 13     | 0.10    | 0.3        | 11 |

<sup>a</sup>Data include at least 3 observations  
N = number of PCR fragments

homozygote type could be a heterozygote on the level of sequence analysis. Fragments of the same length can have different sequence structures, especially in HumVWFA31, which consists of different repeat motifs [1, 20]. Additionally, inbreeding is a relatively likely assumption for observing excess homozygosity in such genetically closed population groups as Hungarian Romanies.

An interclass correlation analysis and an  $s_k^2$  criterion demonstrated that there was no evidence for allelic association between/among the STR loci ( $P \geq 0.064$ ;  $s_k^2 = 0.967$ , 95% C.I. of variance is 0.628–0.986).

A measure of the precision of the computerized allele sizing was examined by analysing the Promega allelic ladders on five different gels. The accuracy values were

**Table 4** G statistic test (*P* values) for homogeneity on VWA, TH01, TPOX, and CSF1PO allele distributions

| Populations | VWA         | TH01        | TPOX        | CSF1PO |
|-------------|-------------|-------------|-------------|--------|
| Romany/CBu  | $< 10^{-3}$ | $< 10^{-3}$ | $< 10^{-3}$ | 0.024  |
| Romany/CBa  | 0.002       | 0.002       | $< 10^{-3}$ | 0.759  |
| CBu/CBa     | 0.184       | 0.678       | 0.458       | 0.668  |

CBu = Hungarian Caucasians residing in the Budapest area [5]  
 CBa = Hungarian Caucasians residing in Baranya county [21, 22]

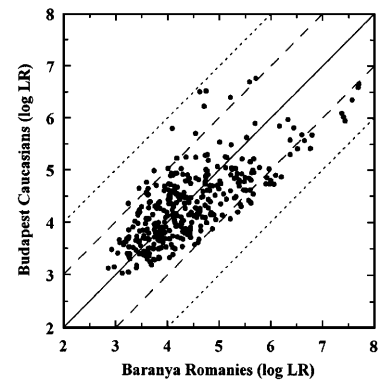
**Table 5** Phenotype frequency differences (mean values) of the STR loci between populations. Populations were compared from two directions. The abbreviation CBu as given in Table 4 A) Single STR systems. B) 2–4 systems combined

| A            |                       |                  |                    |        |
|--------------|-----------------------|------------------|--------------------|--------|
| Populations  | VWA                   | TH01             | TPOX               | CSF1PO |
| Romany → CBu | 1.5                   | 1.4              | 1.5                | 1.4    |
| CBu → Romany | 1.5                   | 1.4              | 1.8                | 1.2    |
| B            |                       |                  |                    |        |
| Population   | 2 loci<br>(VWA, TH01) | 3 loci<br>(TPOX) | 4 loci<br>(CSF1PO) |        |
| Romany → CBu | 2.1                   | 3.2              | 4.6                |        |
| CBu → Romany | 2.1                   | 2.8              | 3.3                |        |

very similar to those data obtained from the analysis of 928 allelic fragments from 135 Romany individuals (Table 3). This extremely high accuracy (99.97% on average for individual samples) allows a clear distinction between the adjacent alleles differing in size by even 1 bp.

The Hungarian Romany allele frequency values for the four STR loci, with the exception of HumCSF1PO, were significantly different from two other Hungarian population databases (Table 4). These two other databases can be considered homogeneous on all the four loci and represent the Hungarian Caucasians residing in the central [5] and the south-western parts [21, 22] of Hungary. The differences in the allele frequency distributions between the Romany and other Hungarian population databases corresponded with previous observations on conventional blood type markers [7, 8]. The combined forensic efficiency values observed in the Romany population sample [ $pM(4 \text{ loci}) = 2.3 \times 10^{-4}$ ;  $MEC(4 \text{ loci}) = 0.93$ ] were similar to those found in the two other Hungarian databases mentioned above.

Interpopulation comparisons with respect to individual phenotypes were carried out between the Baranya Romany and Central Hungarian population databases. The comparison regarding MVs revealed quite small interpopulation differences, which did not increase considerably when the STR loci were combined (Table 5). Thus the investigated STR systems have low discrimination power for the two populations analysed here. Our results are in good accordance with those previous findings, which suggest that generally only minor phenotype frequency dif-

**Fig. 1** A scatter plot comparison of the Baranya Romany and Budapest Caucasian population databases for 358 four-locus STR profiles. The solid diagonal line indicates the theoretical line where both databases would produce the same estimates. The dashed and dotted lines assign the one and two order magnitude differences in the profile estimates, respectively

ferences exist within major population groups [18, 19]. In the scatter plot comparison (Fig. 1) 5.3% of the four-locus profiles had LR estimates differing by more than one order of magnitude where estimates were more common than 1/1 million (6 in log). This fact indicates that in certain cases there may be an unfair bias in applying of the Baranya Romany database to the four-locus STR profiles of the Central Hungarian population and vice versa.

In conclusion, a Hungarian Romany population database has been established for four STR loci using a semi-automatic DNA typing method. Because of its high precision and individualization power, the present STR profiling system is a valuable tool for forensic DNA work in Hungary.

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